



中华人民共和国出入境检验检疫行业标准

SN/T 1950—2007

进出口茶叶中多种有机磷农药 残留量的检测方法 气相色谱法

Determination of organophosphorus pesticides multiresidues
in tea for import and export—Gas chromatography

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中 华 人 民 共 和 国
国家质量监督检验检疫总局 发 布

前 言

本标准的附录 A 为资料性附录。

本标准由国家认证认可监督管理委员会提出并归口。

本标准起草单位：中华人民共和国湖南出入境检验检疫局。

本标准主要起草人：张莹、颜鸿飞、孙菲、黄志强、王美玲、李拥军。

本标准系首次发布的出入境检验检疫行业标准。

进出口茶叶中多种有机磷农药 残留量的检测方法 气相色谱法

1 范围

本标准规定了茶叶中敌敌畏、甲胺磷、乙酰甲胺磷、甲拌磷、氧乐果、乙拌磷、异稻瘟净、乐果、皮蝇磷、毒死蜱、杀螟硫磷、对硫磷、水胺硫磷、杀扑磷、乙硫磷、三唑磷、芬硫磷、苯硫磷、亚胺硫磷、伏杀硫磷、吡啶磷等 21 种有机磷农药残留量的气相色谱测定方法。

本标准适用于茶叶中 21 种有机磷农药残留量的测定。

2 方法提要

试样经水浸泡后,用乙酸乙酯和乙酸乙酯+正己烷(1+1,体积比)溶液提取,过活性炭柱净化,用配备火焰光度检测器的气相色谱仪进行测定,外标法定量。

3 试剂和材料

除特殊规定外,所有试剂均为分析纯,水为蒸馏水。

3.1 乙酸乙酯:重蒸馏。

3.2 正己烷:重蒸馏。

3.3 丙酮:重蒸馏。

3.4 无水硫酸钠:650℃灼烧 4 h。

3.5 敌敌畏、甲胺磷、乙酰甲胺磷、甲拌磷、氧乐果、乙拌磷、异稻瘟净、乐果、皮蝇磷、毒死蜱、杀螟硫磷、对硫磷、水胺硫磷、杀扑磷、乙硫磷、三唑磷、芬硫磷、苯硫磷、亚胺硫磷、伏杀硫磷、吡啶磷等 21 种农药标准品:纯度大于等于 98%。

3.6 农药标准溶液:准确称取适量的单个有机磷农药标准品,用丙酮配成 100 $\mu\text{g}/\text{mL}$ 的储备液,使用时根据需要用乙酸乙酯稀释成适当浓度的混合标准工作液。

3.7 活性碳固相萃取柱:3 mL 活性炭柱(SUPELCO 或相当者)。

4 仪器和设备

4.1 气相色谱仪:配有火焰光度检测器(FPD),磷滤光片(526 nm)。

4.2 快速混匀器。

4.3 离心机:3 000 r/min。

4.4 多功能微量样品处理仪或其他相当的仪器。

4.5 具塞刻度离心管:5 mL、10 mL。

4.6 玻璃试管:10 mL。

4.7 尖嘴吸管。

5 试样制备与保存

5.1 试样制备

取有代表性样品 500 g,用粉碎机粉碎并通过 2.0 mm 圆孔筛。混匀,均分成两份作为试样,分装入

洁净的盛样容器内,密封并标明标记。

5.2 试样保存

将试样于 0℃~4℃ 保存。

在制样的操作过程中,应防止样品受到污染或发生残留物含量的变化。

6 分析步骤

6.1 提取

称取 0.5 g(精确至 0.001 g)试样于 10 mL 试管中,加入 1 mL~1.5 mL 水,浸泡 10 min。加入无水硫酸钠使之饱和后,用 2×2 mL 乙酸乙酯提取两次,每次振荡 2 min,于 2 000 r/min 离心 3 min,收集上层有机相;残渣再用 2 mL 乙酸乙酯-正己烷(1+1,体积比)提取一次,合并上层有机相,待净化。

6.2 净化

在活性碳固相萃取柱上端装入 1 cm 高无水硫酸钠,用乙酸乙酯 4 mL 预淋洗小柱,弃去流出液,然后将提取液全部倾入柱中,再分别用 4 mL 乙酸乙酯和 2 mL 乙酸乙酯+正己烷(1+1,体积比)洗脱,收集全部流出液于 5 mL 具塞刻度离心管中,于 40℃ 下用氮气(流吹至 0.50 mL,供气相色谱分析。

6.3 测定

6.3.1 气相色谱条件

- a) 色谱柱: EQUITY-1701 石英毛细管柱, 30 m×0.53 mm(内径)×1.0 μm, 或相当者;
- b) 升温程序: 100℃(1 min) $\xrightarrow{10^{\circ}\text{C}/\text{min}}$ 160℃(1 min) $\xrightarrow{5^{\circ}\text{C}/\text{min}}$ 240℃(8 min);
- c) 进样口温度: 250℃;
- d) 检测器温度: 250℃;
- e) 载气: 氮气, 纯度大于等于 99.99%, 流量 5.0 mL/min;
- f) 氢气: 75 mL/min;
- g) 空气: 100 mL/min;
- h) 尾吹气: 20 mL/min;
- i) 进样方式: 无分流进样, 1.0 min 后开阀;
- j) 进样量: 2 μL。

6.3.2 色谱测定

根据样液中有机磷含量情况,选定与样液浓度相近的标准工作溶液。标准工作溶液和样液中各种有机磷农药响应值均应在仪器检测线性范围内,标准工作溶液和样液等体积穿插进样测定。在上述气相色谱条件下,参考保留时间为:敌敌畏 4.0 min、甲胺磷 5.1 min、乙酰甲胺磷 7.9 min、甲拌磷 9.2 min、氧乐果 10.2 min、乙拌磷 11.2 min、异稻瘟净 11.9 min、乐果 12.4 min、皮蝇磷 12.8 min、毒死蜱 14.0 min、杀螟硫磷 14.8 min、对硫磷 15.5 min、水胺硫磷 16.1 min、杀扑磷 17.3 min、乙硫磷 19.6 min、三唑磷 21.3 min、芬硫磷 22.3 min、苯硫磷 23.1 min、亚胺硫磷 23.5 min、伏杀硫磷 24.6 min、吡啶磷 25.5 min。

标准品的色谱分离图参见附录 A 中图 A.1。

6.4 空白试验

除不加试样外,按上述测定步骤进行。

7 结果计算和表述

用色谱数据处理机或按式(1)计算试样中有机磷残留量,计算结果需扣除空白值。

$$X = \frac{A \cdot c \cdot V}{A_s \cdot m} \dots\dots\dots(1)$$

式中：

X——样品中有机磷含量,单位为毫克每千克(mg/kg)；

A——样液中有机磷的峰面积；

A_s——标准工作溶液中有机磷的峰面积；

c——标准工作溶液中有机磷的浓度,单位为微克每毫升(μg/mL)；

V——样液最终定容体积,单位为毫升(mL)；

m——称取的试样质量,单位为克(g)。

8 方法的测定低限、回收率

本方法的测定低限和回收率数据见表1。

表1 测定低限和回收率数据

农药名称	添加范围/(mg/kg)	回收率范围/%	测定低限/(mg/kg)
敌敌畏	0.02~0.2	84.0~105	0.02
甲胺磷	0.02~0.2	79.5~122	0.02
乙酰甲胺磷	0.02~0.2	79.0~118	0.02
甲拌磷	0.02~0.2	70.0~94.0	0.02
氧乐果	0.02~0.2	73.5~95.1	0.02
乙拌磷	0.02~0.2	69.8~101	0.02
异稻瘟净	0.01~0.1	82.0~123	0.01
乐果	0.02~0.2	88.9~113	0.02
皮蝇磷	0.02~0.2	75.4~117	0.02
毒死蜱	0.02~0.2	71.9~104	0.02
杀螟硫磷	0.02~0.2	76.6~111	0.02
对硫磷	0.02~0.2	79.7~110	0.02
水胺硫磷	0.01~0.1	86.3~125	0.01
杀扑磷	0.02~0.2	77.7~115	0.02
乙硫磷	0.02~0.2	84.2~119	0.02
三唑磷	0.02~0.2	78.4~109	0.02
芬硫磷	0.01~0.1	78.5~125	0.01
苯硫磷	0.01~0.1	74.8~98.6	0.01
亚胺硫磷	0.02~0.2	77.2~115	0.02
伏杀硫磷	0.02~0.2	85.0~119	0.02
吡啶磷	0.01~0.1	83.5~119	0.02

附 录 A
(资料性附录)
21 种有机磷农药的标准气相色谱图

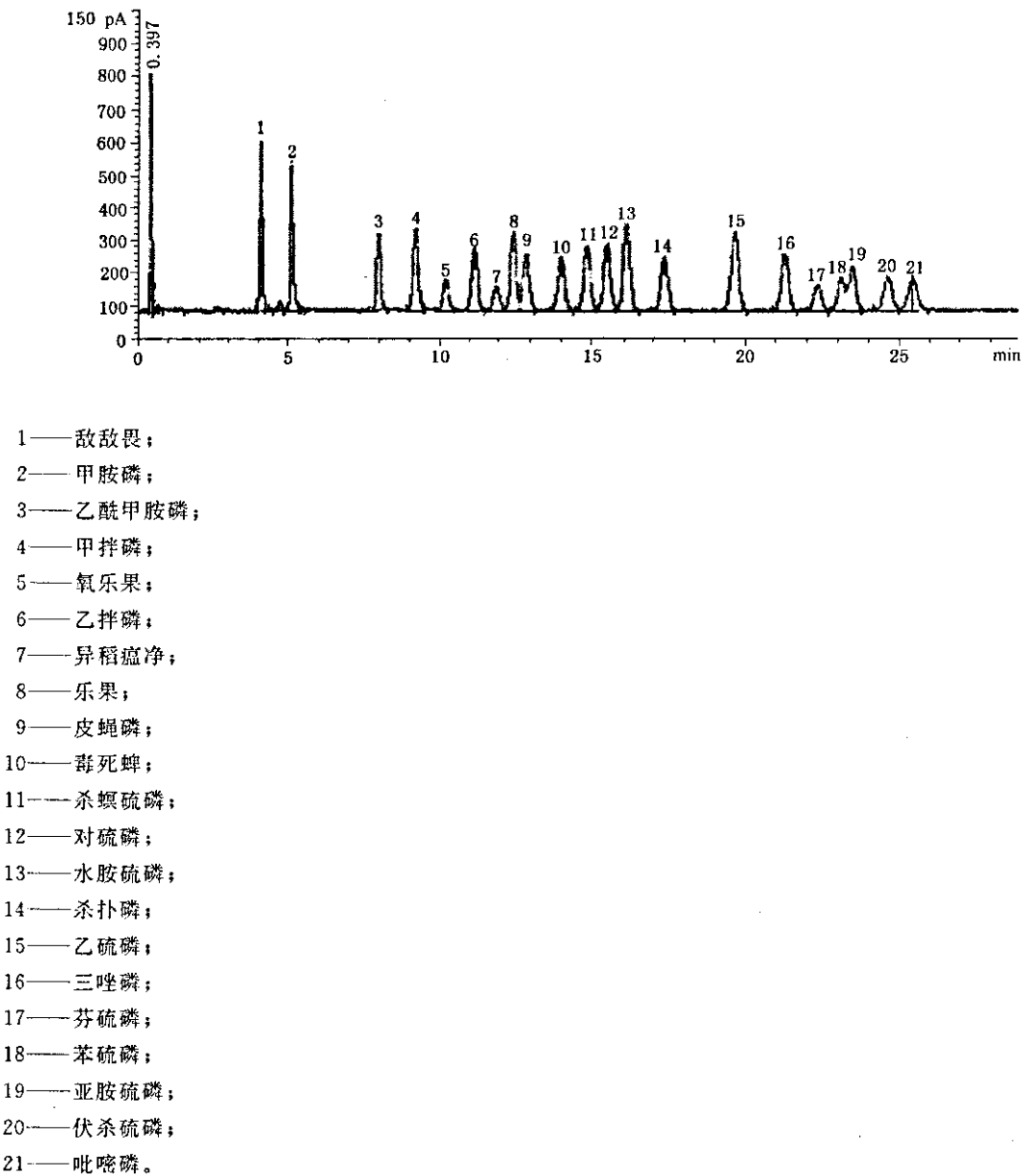


图 A.1 21 种有机磷农药标准品的气相色谱图

Foreword

Annex A of this standard is an informative annex.

This standard was proposed by and is under the charge of the Certification and Accreditation Administration of the People's Republic of China

This standard was drafted by Hunan Entry-Exit Inspection and Quarantine Bureau of the People's Republic of China

The main drafters of this standard are Zhang Ying Yan Hongfei Huang Zhiqiang Wang Meiling Li Yongjun.

This standard is a professional standard for entry-exit inspection and quarantine promulgated for the first time

This standard specifies the method for the determination of dichlorvos, methodophos acephate, phorate omethoate disulfoton IBP dimethoate fenchlorphos chlorpyrifos fenitrothion parathion isocarbophos methidathion ethion triazophos fenstionate EPN phosmet phosalone pyrazophos residues by gas chromatography in tea for import and export.

3 Reagents and materials

Unless otherwise specified all reagents used were of analytical reagent grade Water is distilled water or deionized water.

- 9

in ethyl acetate.

3.7 Envi-Carb column 3 mL(supelco or equivalent).

4 Apparatus and equipment

4.1 Gas chromatograph equipped with a flame photometric detector FPD with a specific filter for phosphorus at 526 nm.

4.2 Vortex mixer.

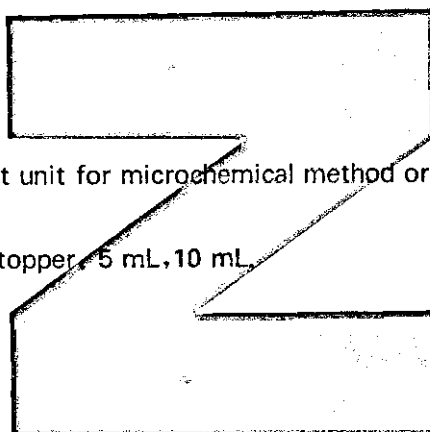
4.3 Centrifuge: 3 000 r/min.

4.4 Multifunction sample treatment unit for microchemical method or equivalent.

4.5 Centrifuge tube with ground stopper: 5 mL, 10 mL.

4.6 Test-tube: 10 mL.

4.7 Glass capillary-tip pipettes.



5 Preparation and storage of test sample

5.1 Preparation of test samples

The combined primary samples is reduced to ca 500 g which is crushed with a grinder and let wholly pass through a 20 mesh sieve. Divided into two equal portions. Each portion is placed in a clean container as the test sample, which is sealed and labeled.

5.2 Storage of test samples

The test samples of tea should be stored below 0℃ ~4℃. In the course of sampling and sample preparation, precaution must be taken to avoid contamination or any factors which may cause the change of residue content.

6 Test Procedure

6.1 Extraction

Weigh 0.5 g of the test sample accurate to 0.001 g in 10 mL centrifuge tube. Marinate the test sam-

ple in 1 mL of water for 10 min. Add enough anhydrous sodium sulfate, and vortex for 2 min. Extract with 2 × 2 mL ethyl acetate by vortex for 2 min, and centrifuge for 3 min. Add 2 mL ethyl acetate-hexane (1 + 1) to the pellet vortex and repeat steps above combining supernatants.

6.2 Cleanup

Add anhydrous sodium sulfate on the Envi-Carb SPE column to the height of 1 cm. Condition the column with 4 mL ethyl acetate. Load the extract solution (6.1) on the column. Elute with 4 mL ethyl acetate and 2 mL ethyl acetate-hexane (1 + 1). Evaporate the eluate to 0.50 mL under nitrogen stream at 40°C. The solution is ready for GC analysis.

6.3 Determination

6.3.1 GC operating conditions

- a) Column: EQUITY-1701, fused silica capillary column 30 m × 0.53 mm (i.d. × 1.0 μm, or equivalent);
- b) Column temperature: 100°C (1 min) $\xrightarrow{10^\circ\text{C}/\text{min}}$ 160°C (1 min) $\xrightarrow{5^\circ\text{C}/\text{min}}$ 240°C (8 min);
- c) Injection port temperature: 250°C;
- d) Detector temperature: 250°C;
- e) Carrier gas: Nitrogen, purity ≥ 99.99%, 5.0 mL/min;
- f) Hydrogen: 75 mL/min;
- g) Oxygen: 100 mL/min;
- h) Make-up gas: 20 mL/min;
- i) Injection mode: Splitless;
- j) Injection volume: 2 μL.

6.3.2 GC determination

According to the approximate concentration of organophosphorus pesticide in the test sample solution, select the standard working solution with similar peak area to that of sample solution. The responses of multiple organophosphorus pesticides in the standard working solution and sample solution should be within the linear range of the instrumental detection. The standard working solution

should be injected randomly in between the injections of sample solution of equal volume Under the above GC opening conditions, chromatogram of organophosphorus pesticide standards are shown in Figure A. 1 in Annex A.

6.3.3 Blank test

The operation of the blank test is the same as that described in the method of determination but with omission of sample addition.

7 Calculation and expression of result

The calculation of organophosphorus pesticide content in the sample is carried out by GC data processor or according to the formula (1), The blank value should be subtracted from the result of calculation

$$X = \frac{A \cdot c \cdot V}{A_s \cdot m} \dots\dots\dots (1)$$

where

- X—the residue content of multiple organophosphorus in the test sample,mg/kg;
- A—the peak area of multiple organophosphorus in the sample solution;
- A_s—the peak area of multiple organophosphorus in the standard working solution;
- c—the concentration of multiple organophosphorus in the standard working solution,μg/mL;
- V—the final volume of sample solution mL;
- m—the corresponding mass of test sample in the final sample solution,g.

8 Limit of determination and recovery

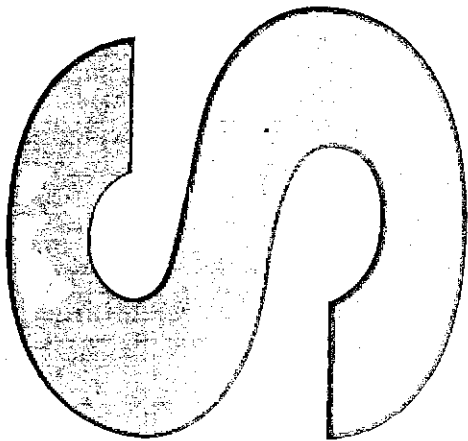
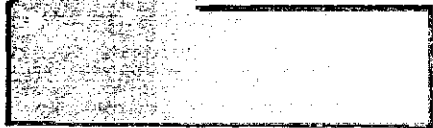
The limit of determination of this method and its corresponding recoveries of fortifying concentrations show on table 1:

Table 1—Limit of determination and recovery

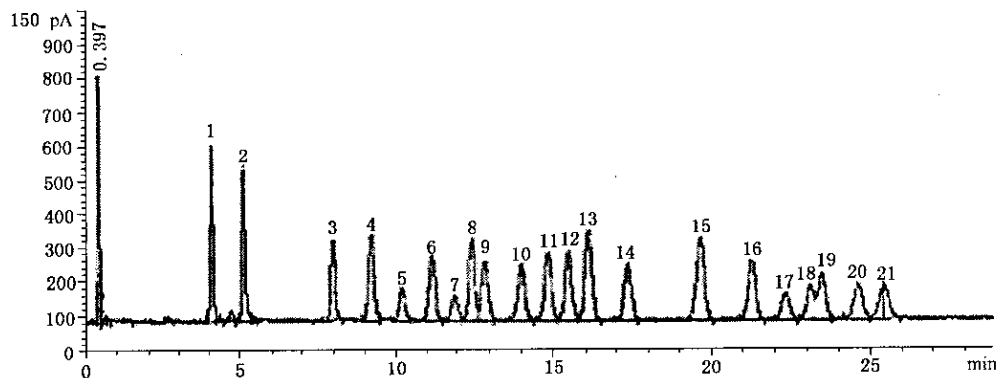
Pesticides	Spiked level/(mg/kg)	Recovery/%	LOD/(mg/kg)
dichlorvos	0.02~0.2	84.0~105	0.02
methodophos	0.02~0.2	79.5~122	0.02
acephate	0.02~0.2	79.0~118	0.02
phorate	0.02~0.2	70.0~94.0	0.02
omethoate	0.02~0.2	73.5~96.1	0.02
disulfoton	0.02~0.2	69.8~101	0.02
IBP	0.01~0.1	82.0~128	0.01
dimethoate	0.02~0.2	88.9~113	0.02
fenchlorphos	0.02~0.2	76.4~117	0.02

Table 1 (continue)

Pesticides	Spiked level/(mg/kg)	Recovery/%	LOD/(mg/kg)
chlorpyrifos	0.02~0.2	71.9~104	0.02
fenitrothion	0.02~0.2	76.6~111	0.02
parathion	0.02~0.2	79.7~110	0.02
isocarbophosl	0.01~0.1	86.3~125	0.01
methidathion	0.02~0.2	77.7~115	0.02
ethion	0.02~0.2	84.2~119	0.02
triazophos	0.02~0.2	78.4~109	0.02
fensthionoate	0.01~0.1	78.5~125	0.01
EPN	0.01~0.1	74.8~98.6	0.01
phosmet	0.02~0.2	77.2~115	0.02
phosalone	0.02~0.2	85.0~119	0.02
pyrazophos	0.01~0.1	83.5~119	0.02



Annex A
(Informative)
Chromatogram of the standards



- 1—Dichlorvos;
- 2—methidophos;
- 3—Acephate;
- 4—Phorate;
- 5—Omethoate;
- 6—Disulfoton;
- 7—IBP;
- 8—Dimethoate;
- 9—Fenchlorphos;
- 10—Chlorpyrifos;
- 11—Fenitrothion;
- 12—Parathion;
- 13—IsocarbophosI;
- 14—Methidathion;
- 15—Ethion;
- 16—Triazophos;
- 17—Fensthiionate;
- 18—EPN;
- 19—Phosmet;
- 20—Phosalone;
- 21—Pyrazophos.

Figure A. 1—Chromatogram of mixed solution of organophosphorus pesticide standards

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